

Phenolic constituents from *Drypetes armoracia*[☆]Jean Wandji^{a,*}, François Tillequin^b, Dulcie A. Mulholland^c, Agathe D. Temgoua^a, Jean-D. Wansi^a, Elisabeth Seguin^d, Z. Tanee Fomum^a^aDepartment of Organic Chemistry, University of Yaoundé-1, Faculty of Science, P.O. Box 812 Yaoundé, Cameroon^bLaboratoire de Pharmacognosie, UMR/CNRS No-8638, Université René- Descartes, Faculté des Sciences Biologiques et Pharmaceutiques, 4-Avenue de l'Observatoire, Paris, France^cDepartment of Chemistry, Natural Products Research Group, University of Natal, Durban, South Africa^dLaboratoire de Pharmacognosie, UFR de Médecine Pharmacie Rouen, 22-Bld Gambetta, 76183 Rouen Cedex 1, France

Received 31 July 2002; received in revised form 1 November 2002

Abstract

The methanol extract of the dried stem bark of *Drypetes armoracia* Pax & Hoffm. afforded two compounds named drypearmoracein A, (E)-4,5,6,7-tetrahydroxy-2-benzylhept-2-enoic acid and drypearmoracein B, 2,3-dihydroxy-9,10-tetrahydroanthra-1,4-quinone along with five known compounds: friedelan-3 β -ol, friedelin, friedelane-3,7-dione, drypemolundein B and β -stigmatsterol. Their structures were established on the basis of spectroscopic analysis and chemical evidence.

© 2003 Elsevier Science Ltd. All rights reserved.

Keywords: *Drypetes armoracia*; Euphorbiaceae; Stem bark; Drypearmoraceins A and B; Hydroxyanthraquinone

1. Introduction

Drypetes armoracia Pax & Hoffm (Euphorbiaceae) is one of more than 20 species of *Drypetes* found in Cameroon. Some of the *Drypetes* species have traditionally been used in West and Central Africa to treat various diseases including sinusitis, boils, swellings, gonorrhoea and dysentery (Dalziel, 1937; Irvine, 1961; Bouquet and Debray, 1974; Walker et al., 1961). Recently, we reported the isolation of a new sesquiterpene lactone and a new friedelane-type triterpene from the stems of *D. molunduan* (Wandji et al., 2000). The total extract and the sesquiterpene lactone were found to exhibit strong anti-inflammatory and analgesic actions (Chungag-Anye et al., 2001, 2002). This background prompted us to explore the chemical constituents of the Cameroonian species, *D. armoracia*. We describe herein the isolation and the structural elucidation, on the basis of extensive NMR studies, of two new compounds: drypearmoracein A, (E)-4,5,6,7-tetrahydroxy-2-benzylhept-2-enoic acid **1** and drypearmoracein B, 2,3-dihydroxy-9,10-tetrahydroanthra-1,4-quinone **2**.

2. Results and discussion

The methanol extract of the air dried stem bark of *D. armoracia* was repeatedly subjected to column chromatography on silica gel to separate compounds **1**–**7**.

Compound **1** was obtained as colourless crystals. The molecular formula of C₁₄H₁₈O₆, containing 6 unsaturation equivalents, was deduced by DEPT, ¹³C NMR, CIMS and FABMS data. Its IR spectrum indicated absorption bands due to the carboxylic acid (1750 cm⁻¹), hydroxyl (3450 cm⁻¹) and double bonds C=C (1600 cm⁻¹) groups. The UV spectrum showed absorption band at λ_{max} 265.2 nm. The ¹H NMR spectrum of **1** indicated a multiplet at δ 7.27–7.34 assigned to the five protons of one monosubstituted benzene ring. It also showed three hydroxymethine protons at δ 4.82 (H-4), 3.51 (H-5) and 3.65 (H-6) and one pair of hydroxymethylene proton signals at δ 3.58 (H-7a) and 3.74 (H-7b). The offset signal at δ 11.82 (*br s*) was attributed to the COOH proton. Thus, these informations suggested that compound **1** contained four hydroxyl groups and one carboxylic acid function. The ¹H NMR spectrum of the acetylated derivative **1a** showed four singlets at δ 2.10, 2.05, 2.02 and 2.00 (3H each), assigned to the acetyl protons, confirming therefore the presence of four hydroxyl groups in **1**. In addition, the ¹H NMR

[☆] Part 4 in the series *Drypetes* studies.^{*} Corresponding author. Tel.: +237-231-09-57; fax: +237-223-53-86.
E-mail address: jeanwandji@yahoo.fr (J. Wandji).

Table 1
¹³C NMR and DEPT data^a and HMBC connectivities of dry-pearmoracein A **1** (100 MHz, DMSO-*d*₆)

Attribution	¹³ C	DEPT	HMBC
1	160.1	C	
2	131.2	C	
3	115.1	CH	C-1, C-2, C-4, C-5, C-7'
4	64.8	CH	C-2, C-3, C-5, C-6
5	73.5	CH	C-3, C-4, C-6, C-7
6	71.4	CH	C-4, C-5, C-7
7	63.6	CH ₂	C-5, C-6
1'	136.7	C	
2'	127.8	CH	C-1', C-3', C-4'
3'	128.5	CH	C-1', C-2', C-4'
4'	127.6	CH	C-2', C-3', C-5' C-6'
5'	128.5	CH	C-1', C-6', C-4'
6'	127.8	CH	C-1', C-5', C-4'
7'	49.8	CH ₂	C-1, C-2, C-3, C-1', C-2', C-6'

^a Assignments were accomplished using ¹H COSY, HETCOR and HSQC experiments.

spectrum of **1** showed one doublet at δ 5.18 ($J=1.20$ Hz), assigned to the CH₂ protons (H-7') of the benzyl group linked to the sp² carbon (C-2) and one broad singlet at δ 6.78 assigned to one olefinic proton (H-3). The decoupled ¹³C NMR and DEPT spectra of **1** (Table 1) showed the presence of 14 carbons. The COOH group carbon at δ 160.1 justified its linkage to the sp² carbon of the double bond. In addition, the ¹³C NMR spectrum showed two double bond carbons at δ 131.1 (C-2) and 115.1 (C-3), six monosubstituted benzene ring carbons at δ 136.7 (C-1'), 128.5 (C-2'/C-6'), 127.8 (C-4') and 127.7 (C-3'/C-5'); the CH₂ group (C-7') was observed at δ 49.8. The presence of four hydroxyl bearing carbons was confirmed by the chemical shifts observed at δ 64.8 (C-4), 73.5 (C-5), 71.4 (C-6) and 63.6 (C-7). Comprehensive analysis of the COSY spectrum revealed many correlations that established the successive couplings between the following protons: H-4 (δ 4.82) and H-5 (δ 3.51), H-5 and H-6 (δ 3.65), H-6 and H-7a/ H-7b (δ 3.58/ 3.74). The cross peak observed between the COOH proton (δ 11.82) and the olefinic proton (δ 6.78) suggested the *trans*-configuration of the double bond C₂=C₃ (see Fig. 1). Furthermore, the

HMBC spectrum (Table 1) showed cross peaks between the COOH carbon (δ 160.1) and H-7' (δ 5.18), H-3 (δ 6.78), confirming that the COOH group was at the *vicinal*-position with the olefinic proton. The stereochemistry of the asymmetric carbons C-4, C-5 and C-6 was not determined. From the complete spectroscopic data and the mass fragments (see experimental), compound **1** was established as (E)-4,5,6,7-tetrahydroxy-2-benzylhept-2-enoic acid, a new constituent named dry-pearmoracein A.

The elemental composition of compound **2**, C₁₄H₁₀O₄ was deduced by the CI/NH₃ MS (m/z 243 [M+H]⁺, 260 [M+NH₄]⁺) and EIMS (m/z 242 [M]⁺), corresponding to 10 insaturation equivalents. The presence of a hydroxyquinonic moiety in **2** was suggested by its UV spectrum data at λ_{max} 267.2, 263.0 and 258.0 nm. This suggestion was further confirmed by the IR spectrum indicating absorption bands due to OH (3400 cm⁻¹), quinonic C=O (1660 cm⁻¹) and double bond C=C (1600 cm⁻¹). The ¹³C NMR spectrum of **2** (Table 2) showed only seven carbon signals. The hydroxyquinonic moiety in **2** was confirmed by the presence of two ketonic functions at δ 184.3 (C-1 and C-4) with a very low intensity and two oxygenated quaternary carbons at δ 158.2 (C-2 and C-3). These data were comparable to those of a methoxyquinonic moiety reported for some quinone derivatives (Guntern et al., 2001). Comprehensive analysis of the seven ¹³C NMR signals suggested the presence of one perfect symmetry in **2**. Therefore, its molecular formula C₁₄H₁₀O₄ contained two identical parts represented as (C₇H₅O₂)₂. Examination of its DEPT spectrum revealed the presence of three signals only, assigned to the protonated carbons at δ 127.4 (CH), 127.6 (CH) and 44.5 (CH₂). In addition, both the HETCOR and HSQC spectra showed three cross peaks only between the protonated carbons and the corresponding protons at δ_{C} 44.5/ δ_{H} 4.42, δ_{C} 127.4/ δ_{H} 7.21 and δ_{C} 127.6/ δ_{H} 7.29. The ¹H NMR spectrum of **2** (Table 2) showed five signals of which four confirmed the presence of the CH₂ group protons as two singlets at δ 4.26 and 4.28 (H-9/H-10) and the CH protons at δ 7.21 (*d*, $J=7.0$ Hz, H-5/H-8) and 7.29 (*d*, $J=7.0$ Hz, H-6/H-7). The fifth signal at δ 5.02 (*br. s*) was deduced to be the

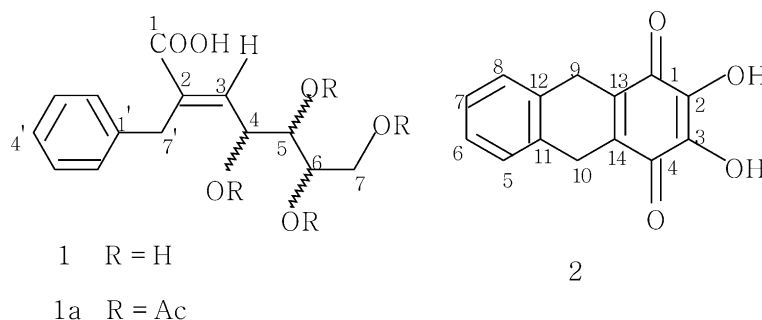


Fig. 1. Structures of compounds **1** and **2**.

Table 2
¹³C, DEPT and ¹H NMR data^a for drypearmoracein **2** in CDCl₃

Position	¹³ C	DEPT	¹ H (J, Hz)
1/4	184.3 ^b	C	
2/3	158.2	C	5.02, <i>br s</i> , OH
5/8	127.4 ^c	CH	7.21, <i>d</i> (7.0)
6/7	127.6 ^c	CH	7.29, <i>d</i> (7.0)
9/10	44.5	CH ₂	4.26; 4.28, <i>s</i> , each
11/12	127.2	C	
13/14	139.1	C	

^a Assignments were accomplished using ¹H COSY, HETCOR and HSQC.

^b Very low intensity.

^c Values can be reversed.

two hydroxyl groups located at the C-2 and C-3 positions in agreement with the presence of the symmetry in **2** (see Fig. 1). At these positions, the two OH groups influence by electronic effects the quinonic ketones at C-1 and C-4, and could then justify the low intensity of their signal at δ 184.3. The complete assignments were accomplished using ¹H–¹H COSY experiment. In accordance to the above spectral data, the arrangement of the elemental formula, in respect to the presence of one perfect symmetry, established the structure of compound **2** as 2,3-dihydroxy-9,10-tetrahydroanthra-1,4-quinone. The structure **2** was supported by its EIMS with some selected important fragments at *m/z* 240 [M–2H] (42), 106 [C₆H₁₀] (100) and 91 [C₇H₇] (80). Compound **2** is new anthra-1,4-quinone derivative named drypearmoracein **B**.

Compounds **3–7** were known constituents isolated for the first time from *D. armoracia*. Their ¹³C NMR spectral data compared to those of the literature identified them as friedelan-3 β -ol **3**, friedelin **4**, friedelane-3,7-dione **5** (Mahato and Kundu, 1994), drypemolundein **B** **6** (Wandji et al., 2000) and β -stigmasterol **7** (Tandon et al., 1990).

3. Experimental

3.1. General

MPs were determined using a Kofler microhot stage apparatus. IR spectroscopy was performed on a Perkin-Elmer 257 spectrometer. $[\alpha]_D$ were read on a Perkin-Elmer 241 polarimeter. UV spectra were recorded on a Beckman DU-600 spectrometer. MS were registered on a Micromass Q-ToF instrument. NMR experiments were performed on a Varian Gemini 400 MHz instrument and a Bruker AC 300 spectrometer. The solvents used for NMR were CDCl₃ and DMSO-*d*₆, also considered in each case as internal references. Si gel 60H (5–40 μ m), 60C (20–40 μ m) and 60 (240–400 mesh) were used for CC under compressed air (300 mbar) while

precoated Si gel 60 F₂₅₄ aluminium plates were used for TLC.

3.2. Plant material

The stem bark of *Drypetes armoracia* Pax and Hoffm was collected from the Dja forest, (East Province of Cameroon) in February 1998. The herbarium specimen documenting the collection has been deposited in the National Herbarium, Yaoundé, Cameroon (Ref. RL.12120).

3.3. Extraction and isolation

Air-dried and powdered stem bark of *Drypetes armoracia* (3.4 kg) was extracted with MeOH (3×4 l) for (3×48 h) at room temperature. After the evaporation of the solvent under reduced pressure, the crude MeOH extract (190.0 g) was obtained. Part of the extract (150.0 g) was chromatographed on Si gel 60 (230–400 mesh) (600 g) using hexane, hexane–EtOAc, EtOAc–MeOH and MeOH in order of increasing polarity. A total of 80 fractions (400 ml each) were eluted. On the basis of the TLC, the fractions were combined into five main series of fractions [A–E]: A (8.6 g) (fractions 5–18) and B (1.3 g) (fractions 19–40), both series eluted with EtOAc–hexane from 25:75 to 30:70; C (5.6 g) (fractions 41–63), eluted with EtOAc–hexane from 40:60 to 100:0; D (12.2 g) (fractions 64–74), eluted with MeOH–EtOAc from 5:95 to 50:50 and E (40.0 g) (fractions 75–80), eluted with MeOH–EtOAc from 60:40 to 100:0. Repeated CC of series A (8.6 g) on Si gel 60C (20–40 μ m) using hexane–EtOAc in increasing percentages yielded compounds **3** (350 mg), **4** (200 mg), **5** (300 mg), **6** (40 mg) and **7** (120 mg). Further CC of series B (1.3 g) on Si gel 60 H (5–40 μ m) using hexane–EtOAc in increasing proportions yielded compound **2** (30 mg). Further CC of series D (12.2 g) on Si gel 60H (5–40 μ m) using EtOAc–MeOH in increasing polarities afforded compound **1** (220 mg).

3.3.1. Drypearmoracein A [(E)-4,5,6,7-tetrahydroxy-2-benzylhept-2-enoic acid] **1**

Colourless crystals; mp 250 °C; $[\alpha]_D +27^\circ$ (MeOH, *c* 0.750); IR (KBr) λ_{\max} (cm^{−1}): 3450 (OH), 1750 (COOH), 1600 (C=C); UV [MeOH] nm (log ϵ): 265.2 (0.7904); CI/NH₃ MS *m/z* 283 [M+H]⁺, 300 [M+NH₄]⁺; FAB MS *m/z* (rel. int.): 289 [M+Li]⁺ (71.2), 283 [M+H]⁺, 280 (27.3), 253 (28.1), 252 (100.0), 246 (47.6), 243 (32.0), 226 (40.6); EIMS, *m/z* 274, 262, 241, 203, 201, 115, 91; ¹H NMR (400 MHz, DMSO-*d*₆): δ 11.82 (1H, *br s*, COOH), 7.27–7.34 (5H, *m*, C₆H₅–), 6.78 (1H, *br s*, H-3), 5.18 (2H, *d*, *J* = 1.20 Hz, H-7'), 4.82 (1H, *br s*, H-4), 3.74 (1H, *dd*, *J* = 2.92, 10.98 Hz, H-7a), 3.65 (1H, *dd*, *J* = 2.65, 8.15 Hz, H-6), 3.58 (1H, *dd*, *J* = 2.92, 10.98 Hz, H-7b), 3.51 (1H, *d*, *J* = 8.15 Hz, H-5); ¹³C

NMR (100 MHz, DMSO- d_6) and HMBC connectivities: see Table 1.

Acetylation of **1**: 10.0 mg of **1** was dissolved in the Ac₂O–pyridine mixture (1:1 ml) and heated at 60 °C for 24 h. After working, the crude product was purified on Si gel 60H to yield one acetylated derivative **1a** (12.5 mg, 78%).

3.3.2. Acetyldrypearmoracein A [(E)-4,5,6,7-tetraacetoxy-2-benzylhept-2-enoic acid] **1a**

Amorphous solid; CI/NH₃ MS, m/z 451 [M+H]⁺; M.F. C₂₂H₂₆O₁₀; ¹H NMR (300 MHz, CDCl₃): δ 11.80 (*br s*, COOH), 7.30–7.35 (5H, *m*, Ar-), 6.85 (*br s*, H-3), 5.20 (*d*, $J=1.20$ Hz, H-7'), 5.12 (*br s*, H-4), 4.04 (*dd*, $J=2.92$, 10.00 Hz, H-7a), 4.02 (*dd*, $J=2.65$, 8.15 Hz, H-6), 4.00 (*dd*, $J=2.92$, 10.00 Hz, H-7b), 3.98 (*d*, $J=8.15$ Hz, H-5), 2.10 (3H, *s* Ac), 2.05 (3H, *s*, Ac), 2.02 (3H, *s*, Ac), 2.00 (3H, *s*, Ac).

3.3.3. Drypearmoracein B [2,3-dihydroxy-9,10-tetrahydroanthra-1,4-quinone] **2**

White crystals; mp 160 °C; IR (KBr) λ_{\max} (cm⁻¹): 3400 (OH), 1660 (C=O), 1600 (C=C); UV (CHCl₃) nm (log ϵ): 267.2 (0.1015), 263.0 (0.1307), 258.0 (0.1667), 252.6 (0.1450), 236.0 (0.0782); EIMS m/z (rel. int.): 242 [M]⁺ (5), 241 [M-H]⁺ (7), 240 [M-2H]⁺ (42), 149 (23), 107 (12), 106 (100), 104 (14), 91 (80), 79 (32), 65 (31), 51 (20); ¹H NMR (300 MHz, CDCl₃) and ¹³C NMR (75 MHz, CDCl₃) values: see Table 2.

Compounds **3**, **4** and **5** were identical to friedela-3 β -ol, friedelin and friedelane-3,7-dione respectively by comparison of their ¹³C NMR data with those reported (Mahato and Kundu, 1994).

Compound **6** was identical to drypemolundein B (friedelane-3-11-dione) previously isolated from *D. molunduana* (Wandji et al., 2000).

Compound **7** was identified to β -stigmasterol by the comparison of its NMR spectral data to the literature values (Tandon et al., 1990).

The friedelane-type triterpenes and sterols represented more than 50% of the constituents isolated from *D. armoracia*.

Acknowledgements

One of the authors (J. Wandji) is grateful for IFS grant No F/2624-2 (Sweden), for the sponsorship of the “Université René- Descartes Paris V, France” during his research visit in Paris, and for the financial support from the OPCW (The Netherlands), through the Internship at the University of Natal, Durban, South Africa. Thanks are due Mr. Dilip Jagjivan for the measurements of some NMR spectra and to Mr. Zacharie D. Nzoo for the identification and the collection of this plant.

References

- Bouquet, A., Debray, L., 1974. Plantes Médicinales de la Côte-d'Ivoire; Travaux et Documents de l'ORSTOM No. 32, pp. 82–87.
- Chungag Anye, N.B., Njamen, D., Dongmo, A.B., Wandji, J., Nguelefack, T.B., Wansi, J.D., et al., 2001. Anti-inflammatory and analgesic properties of the stem extract of *Drypetes molunduana* Pax and Hoffm. (Euphorbiaceae) in rats. *Pharmaceutical and Pharmacological Letters* 11, 61–63.
- Chungag-Anye, N.B., Njamen, D., Wandji, J., Fomum, T.Z., Dongmo, A., Nguelefack, T.B., et al., 2002. Anti-inflammatory and analgesic effects of Drypemolundein A, a sesquiterpene lactone from *Drypetes molunduana*. *Pharmaceutical Biology* 40 (in press).
- Dalziel, J.M., 1937. The Useful Plants of West Tropical Africa. The Crown Agents for the Colonies, London.
- Guntern, A., Ioset, J.-R., Queiroz, E.F., Foggini, C.M., Hostettmann, K., 2001. Quinones from *Heliotropium ovalifolium*. *Phytochemistry* 58, 631–635.
- Irvine, R.F., 1961. Woody Plant of Ghana. Oxford University Press, London.
- Mahato, S.B., Kundu, A.P., 1994. ¹³C NMR spectra of pentacyclic triterpenoids. A compilation and some salient features. *Phytochemistry* 37, 1517–1575.
- Tandon, M., Shukla, Y.N., Thakur, R.S., 1990. Steroid glycosides from *Asparagus adscendens*. *Phytochemistry* 29, 2957–2959.
- Walker, A.R., Sillans, R., Trochain, J.L., 1961. Les Plantes Utiles du Gabon. Ed. Paul Lechevalier 12-Rue de Tournon Paris VI, pp. 165–166.
- Wandji, J., Wansi, J.D., Fuendjie, V., Dagne, E., Mulholland, D.A., Tillequin, F., et al., 2000. Sesquiterpene lactone and friedelane derivative from *Drypetes molunduana*. *Phytochemistry* 54, 811–815.